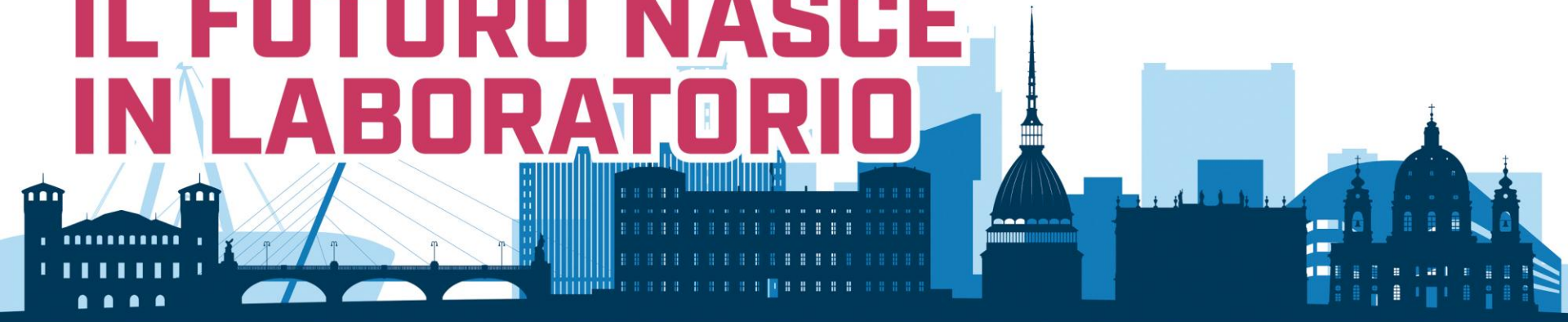


YOUNG SCIENCE FORUM: IL FUTURO NASCE IN LABORATORIO

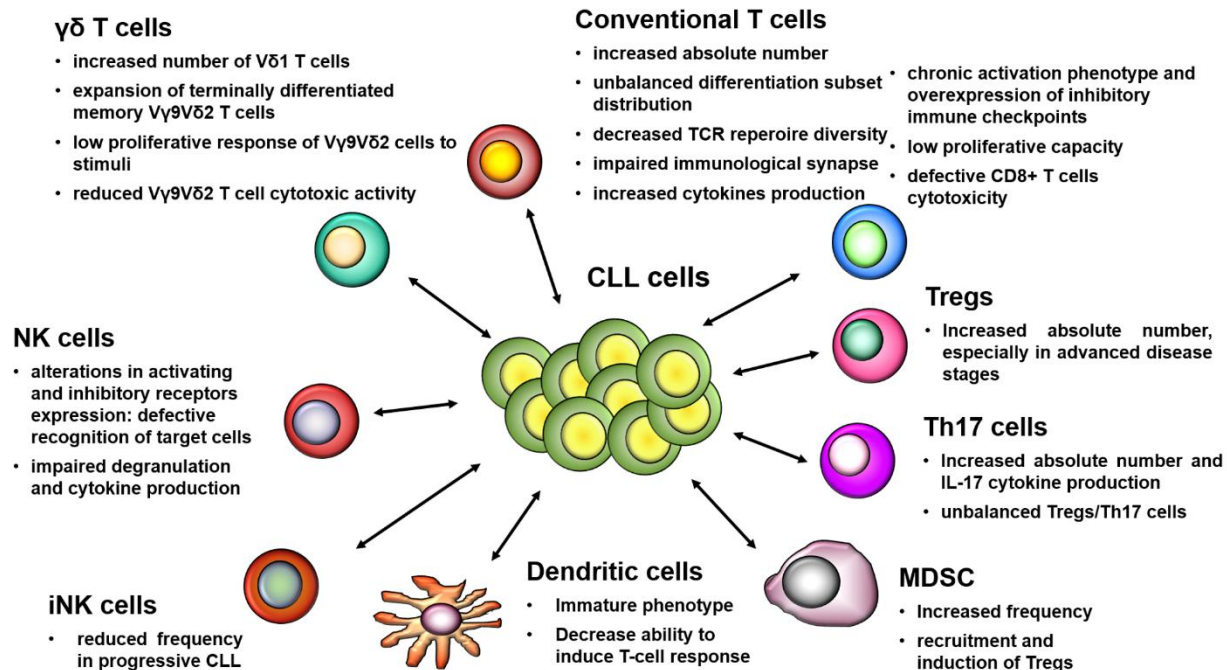


**Setup of an in-house small-scale anti-CD19 CAR T-cell manufacturing process:
immunophenotypical characterization and functional assessments**

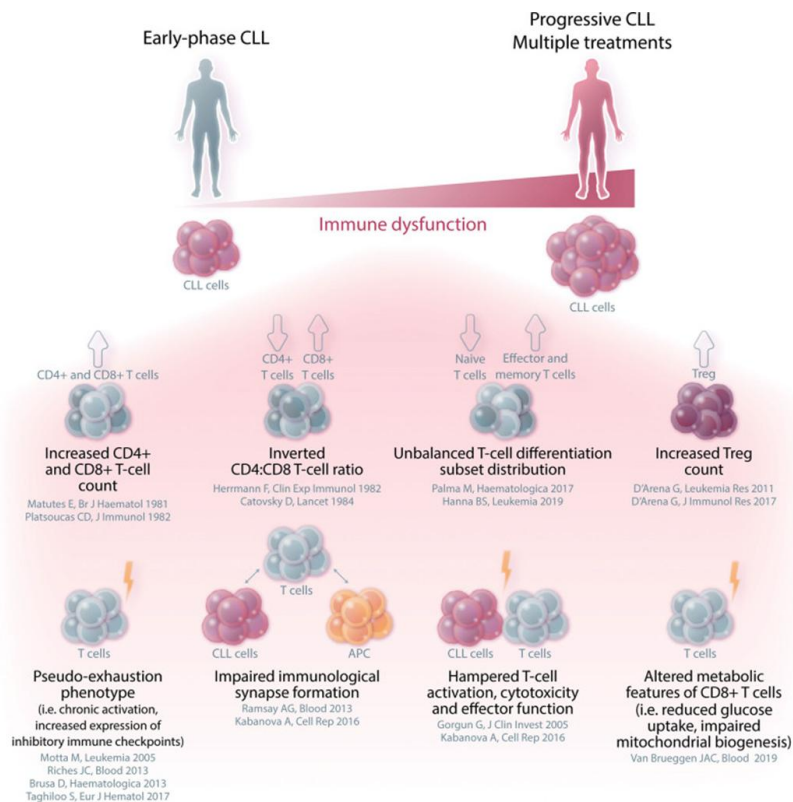
Valentina Griggio

*Laboratory of Translational Hematology - Hematology Section
Department of Molecular Biotechnologies and Health Sciences
University of Torino*

TORINO, ACCADEMIA DI MEDICINA | 4-5 GIUGNO 2026



The complex interactions between **leukemic cells**, **microenvironment** and **immune system** in CLL



T-cell compartment, in particular, is characterized by multiple phenotypical and functional alterations

The **clinical correlates** related to immune dysfunction in CLL

- infections
- autoimmune phenomena
- second primary malignancies
- poor response to vaccines
- suboptimal efficacy of immunotherapy

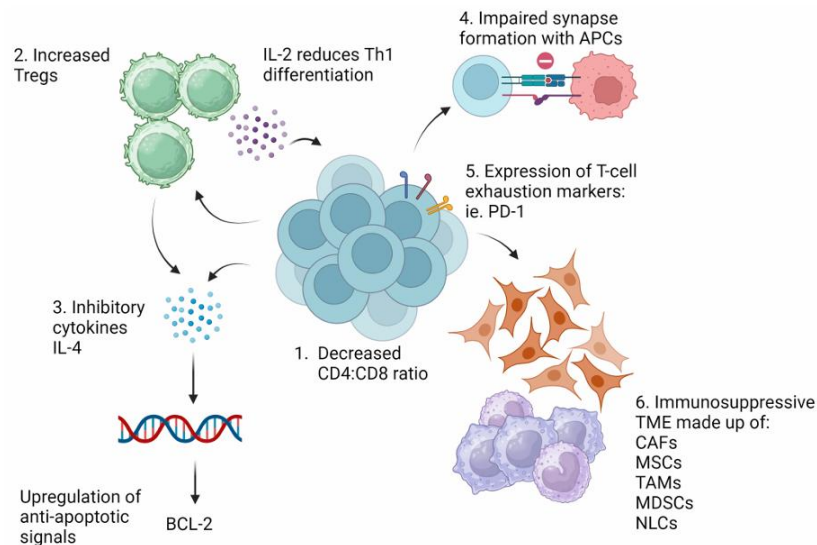
While **CD19 CAR T-cell therapy** has revolutionized the treatment of B-cell malignancies, its efficacy in CLL remains suboptimal:

Clinical trial involving limited number of participants with CLL

Less deep and durable responses compared with other B-cell malignancies

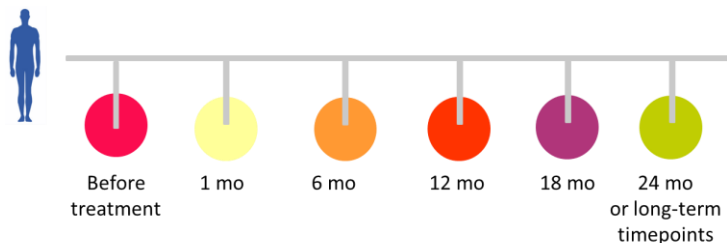
Product Administered	Product Specifics	Patients Evaluable for Response, Number	Lymphodepleting Chemotherapy	Efficacy			Safety			Reference
				ORR	CR Rate	Median DOR	Median PFS	Median OS	Safety	
Axi-cel (KTE-C19) + M IL-2	Second generation	CLL = 4	Flu/Cy	75% (3/4)	25% (1/4)	7 months	NA	NA	Grade ≥3 AEs 100% (4/4)	67
Axi-cel (KTE-C19)	Second generation	CLL = 4 RS = 1	Flu/Cy	CLL: 100% (4/4) RS: 100% (0/1)	CLL: 75% (3/4) RS: 0% (0/1)	CLL: 14 months RS: 1 months	NA	NA	CLL: grade ≥3 AEs 50% (2/4) RS: grade ≥3 AEs 100% (1/1)	68
Brexi-cel (KTE-X19)	Second generation; depletion of circulating CD19-expressing malignant cells	CLL = 15	Flu/Cy	47% (7/15)	13% (2/15)	NA	NA	NA	Grade ≥3 AEs 100% (15/15) CRS 80% (12/15), grade ≥3 CRS 7% (1/15) NE 73% (11/15), grade ≥3 NE 20% (3/15)	70
Tisa-cel (CTL019)	Second generation	CLL = 3	Different regimens (bendamustine ± rituximab, pentostatin/Cy)	100% (3/3)	33% (1/3)	10 months	NA	NA	NA	71
Tisa-cel (CTL019)	Second generation	CLL = 14*	Different regimens (Flu/Cy, bendamustine, pentostatin)	57% (8/14)	29% (4/14)	CR patients, 40 months PR patients, 7 months	7 months	29 months	CRS 63% (9/14), grade ≥3 CRS 43% (6/14) NE 36% (5/14), grade ≥3 NE 7% (1/14)	72
Tisa-cel (CTL019)	Second generation	CLL = 32 (5 × 10 ⁶ CAR T cells, HD = 19; 5 × 10 ⁶ CAR T cells, LD = 13)	Different regimens (bendamustine, Flu/Cy, pentostatin/Cy, oxaliplatin/Flu/cytarabine/rituximab, gemtacin/oxaliplatin)	HD: 53% (10/19) LD: 31% (4/13)	HD: 37% (7/19) LD: 15% (2/13)	NA	1 months*	64 months*	CRS 63% (24/38), grade ≥3 CRS 24% (9/38)* Grade ≥3 NE 8% (3/38)*	73
MSKCC CD19 CAR T cells	Second generation	CLL = 8	None or Cy	0%	0%	NA	NA	NA	Grade ≥3 AEs 50% (4/8)	75
MSKCC CD19 CAR T cells	Second generation	CLL = 16*	None or different regimens (Cy, bendamustine, Flu/Cy)	38% (6/16)	19% (3/16)	NA	3 months	17 months	CRS 100% (16/16), grade ≥3 CRS 12% (2/16) NE 38% (6/16), grade ≥3 NE 6% (1/16) CLL: CRS 95% (18/19), grade ≥3 CRS 10% (2/19), NE 37% (7/19), grade ≥3 NE 26% (5/19) RS: CRS 40% (2/5), grade ≥3 CRS 0, NE 20% (1/5), grade ≥3 NE 20% (1/5) CRS 85% (9/11), grade ≥3 CRS 10% (1/11) NE 45% (5/11), grade ≥3 NE 19% (2/11)	76
JCAR014	Second generation; 1:1 ratio of CD4+CD8+ CAR T cells; ex vivo stimulation with antigen-presenting cells	CLL = 18 RS = 5	Cy, Flu, or Flu/Cy	CLL: 72% (13/18) RS: 60% (3/5)	CLL: 11% (2/18) RS: 40% (2/5)	NA	8.5 months*	NR	CRS 100% (18/18), grade ≥3 CRS 10% (2/18) NE 37% (7/19), grade ≥3 NE 26% (5/19) RS: CRS 40% (2/5), grade ≥3 CRS 0, NE 20% (1/5), grade ≥3 NE 20% (1/5) CRS 85% (9/11), grade ≥3 CRS 10% (1/11) NE 45% (5/11), grade ≥3 NE 19% (2/11)	77
Liso-cel (JCAR017)	Second generation; 1:1 ratio of CD4+CD8+ CAR T cells; removal of non-T-cell impurities before activation and transduction; ex vivo stimulation with cytokines	CLL = 96	Flu/Cy	48% (46/96)	18% (17/96)	35 months (CR/CR)	18 months	NA	CRS 85% (99/117), grade ≥3 CRS 10% (10/117) NE 45% (53/117), grade ≥3 NE 19% (22/117)	88
CD19 CAR T cells (Baylor College of Medicine)	Third generation with 2 costimulatory domains (CD28 and 4-1BB)	CLL = 2	Flu/Cy	50% (1/2)	0%	NA	13 months	>40 months	CRS 2% (3/15), grade ≥3 CRS 7% (1/15)* NE grade ≥3 13% (2/15)*	89
CD19 CAR T cells (Heidelberg University Hospital)	Third generation with 2 costimulatory domains (CD28 and 4-1BB)	CLL = 4	Flu/Cy	43%*	29%*	NA	NA	NA	CRS grade ≥3 0.7% (2/27)* NE grade ≥3 0%*	90
MSKCC CD19 CAR T cells	Fourth generation with 2 signaling domains (CD28 and CD3ζ) and 4-1BB ligand expression	CLL = 9 RS = 3	Flu/Cy or Cy alone	CLL: 44% (4/9) RS: 67% (2/3)	CLL: 33% (3/9) RS: 67% (2/3)	NA	NA	NA	CRS 39% (11/28), grade ≥3 CRS 3% (1/28)* CRS 39% (11/28), grade ≥3 CRS 10% (3/28)*	91
JCAR014 and brutinib	Second generation	CLL = 18	Flu/Cy	83% (15/18)	22% (4/18, all CR)	NA	NA (1-year PFS, 38%)	NA (1-year OS, 64%)	CRS 74% (14/19), grade ≥3 CRS 0% NE 26% (5/19), all grade 3	112
CTL119 and brutinib	Second generation	CLL = 16	Flu/Cy or bendamustine	69% (11/16)	44% (7/16)	NA	NR (48-months PFS, 70%)	NR (48-months OS, 84%)	CRS 35% (18/19), grade ≥3 CRS 11% (2/19) NE 26% (5/19), grade ≥3 NE 5% (1/19)	113
Liso-cel (JCAR017) and brutinib	Second generation	CLL = 19	NA	95% (19/19)	47% (9/19) CR/CR	NA	NA	NA	CRS 74% (14/19), grade ≥3 CRS 5% (1/19) NE 32% (6/19), grade ≥3 NE 16% (3/19)	114

One of the major determinants of CAR T-cell therapy limited efficiency is T-cell exhaustion and a tumor-supportive TME



Therefore, a deeper understanding of these mechanisms is essential to improve CAR T-cell manufacturing and therapeutic outcomes

One of our laboratory's main research interest is the study of the **immunomodulatory properties** of targeted therapies. Besides their direct anti-tumor activity, targeted agents such as **ibrutinib** or **venetoclax** have shown positive immune changes



- Immunophenotype
- Functional assays
- Correlation with prognostic factors, disease and response variables

Immune changes are possibly linked to **off-tumor and off-target effects** and could also be associated to the reduction of tumor burden

641. CLL: BIOLOGY AND PATHOPHYSIOLOGY, EXCLUDING THERAPY: POSTER III | NOVEMBER 29, 2018

Ibrutinib Treatment Mitigates Phenotypic Alterations of Non-Neoplastic Immune Cell Compartments in Chronic Lymphocytic Leukemia

Valentina Griggio, Candida Vitale, MD, Maria Todaro, PhD, Francesca Romana Mauro, Chiara Salvetti, MD, Daniela Pietrasanta, Iolanda Donatella Vincelli, MD, Lydia Scarfo, MD, Giovanni Del Poeta, MD, Gianluca Gaidano, MD PhD, Valter Gattei, MD, Robin Foà, MD, Mario Boccadoro, MD, Marta Coscia, MD



Blood (2018) 132 (Supplement 1): 4412.

<https://doi.org/10.1182/blood-2018-99-116068>

641. CHRONIC LYMPHOCYTIC LEUKEMIA: BASIC AND TRANSLATIONAL | NOVEMBER 3, 2025

Venetoclax-based therapy remodels T-cell and NK-cell immunity in patients with chronic lymphocytic leukemia

Giorgia Mancin, Valentina Griggio, Rebecca Jones, Giulia Bondielli, Andrea Visentin, Riccardo Moia, Anna Maria Frustaci, Alessandro Bosi, Marzia Varettoni, Francesca Perutelli, Maria Chiara Montalbano, Gioacchino Catania, Paolo Sportoletti, Luca Laurenti, Francesca Mauro, Lorella Orsucci, Luca Arcaini, Francesco Passamonti, Marina Deodato, Gianluca Gaidano, Livio Trentin, Benedetto Bruno, Candida Vitale, Marta Coscia



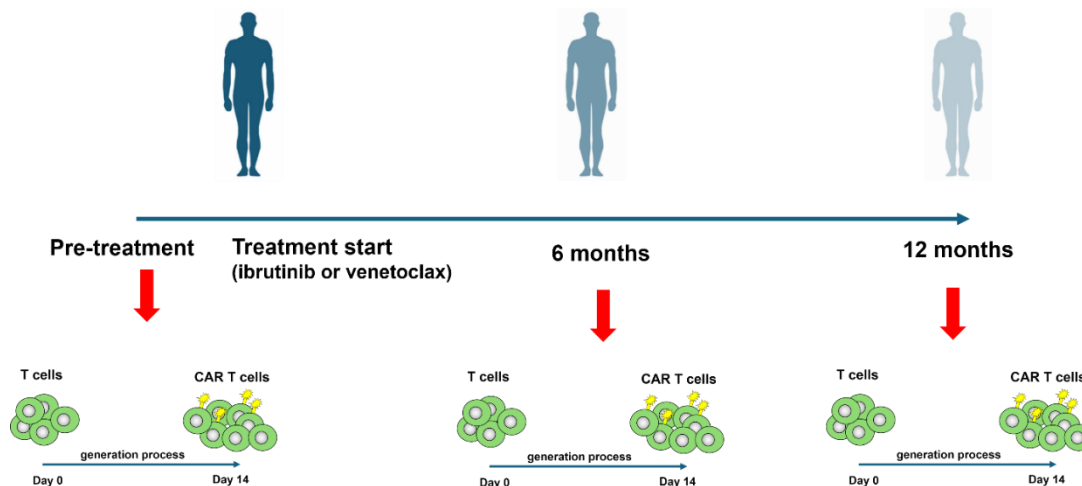
Blood (2025) 146 (Supplement 1): 5657.

<https://doi.org/10.1182/blood-2025-5657>

The **GIMEMA CLL2020** study aims to evaluate, in CLL patients, the impact of treatment with targeted agents (i.e. **ibrutinib** or **venetoclax**) on T cells and on patient-derived **anti-CD19 CAR T** cells in terms of:

- manufacturing efficiency
- immunophenotypical profile
- functional properties

Samples were collected before the start of treatment and after 6 and 12 months of therapy with ibrutinib (n=7) or venetoclax (n=8)



The functionality and persistence of infused CAR T cells are strictly connected to the composition and the fitness of both the starting T-cell source and the final CAR T-cell product

CD19 CAR T-cell generation workflow

Starting material – day 0

PBMC usually contain a large number of leukemic B cells

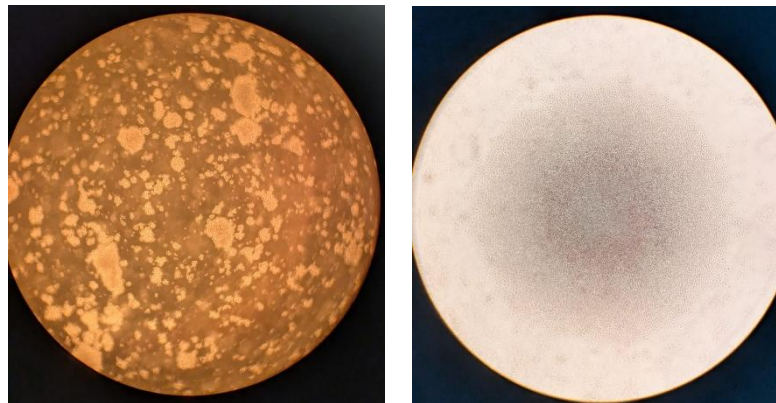
→ B-CELL DEPLETION (positive selection of leukemic cells)

→ T-CELL ENRICHMENT (negative selection of T cells)

CD19 CAR T-cell generation workflow

T-cell activation – day 0

Delayed activation or fail to activate when working with thawed PBMC



→ T CELLS ARE ACTIVATED USING A COLLOIDAL MATRIX CONTAINING α CD3 and α CD28 agonists

→ Early T-cell activation is evaluated by the appearance of characteristic cellular clusters under light microscopy. When sufficient material is available, activation status is further characterized by flow cytometry through the expression of **CD69** and **CD25**

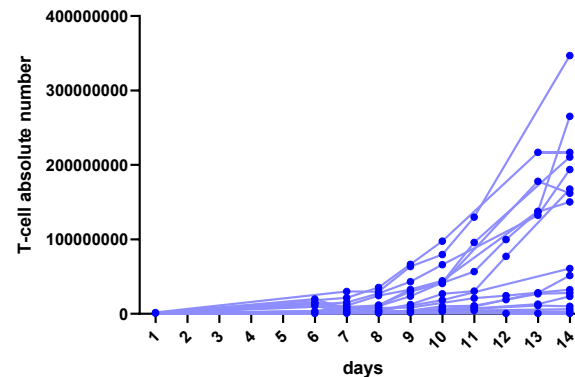
Our **CD19 CAR T-cell generation workflow**

CAR T-cell expansion – approx 9 days

IL-7/IL-15-driven expansion depends on:

- T-cell differentiation status
- T-cell exhaustion
- metabolic fitness

T-cell proliferative ability can vary substantially among CLL patients depending on disease-related immune dysfunction and baseline T-cell composition



→ CAR T CELLS ARE CULTURED IN THE PRESENCE OF IL-7 AND IL-15

CD19 CAR T-cell immunophenotypical characterization

Immune determinants of response to CAR T-cell therapy

Favourable

- Memory-like T-cell phenotype
- High ex vivo proliferative potential
- IL-6/STAT3 pathway activity

Unfavourable

- Higher PD-1, TIM-3 and LAG-3 expression
- Terminal differentiation

Fraietta JA et al., Nature Medicine (2018)

Lineage composition
CD3, CD4, CD8

T-cell differentiation subsets
CCR7, CD45RA, CD95

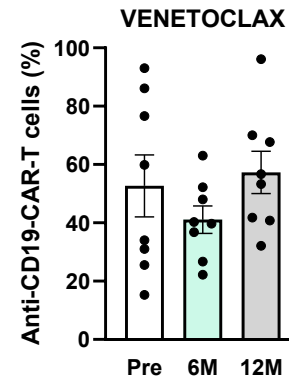
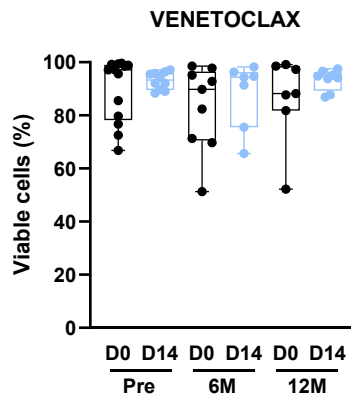
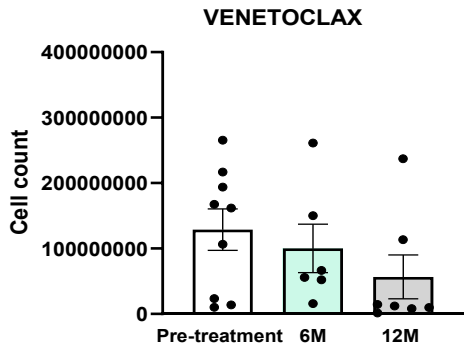
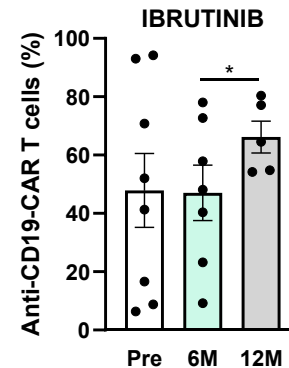
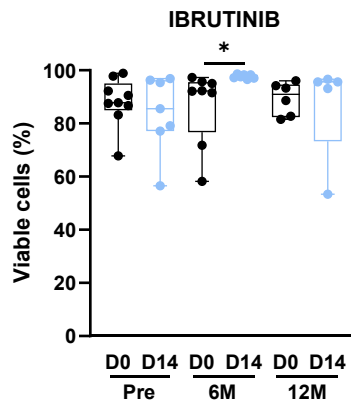
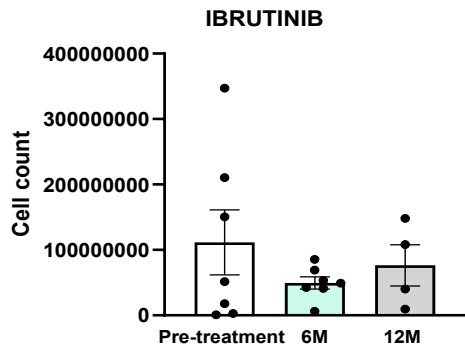
Activation markers
CD25, CD69, HLA-DR

T helper subsets
CCR6, CXCR3

Immune checkpoint
PD-1, TIM-3, CTLA-4,
TIGIT, BTLA

Migration markers
CD62L, CXCR3

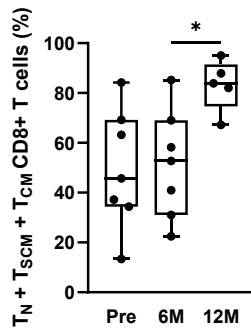
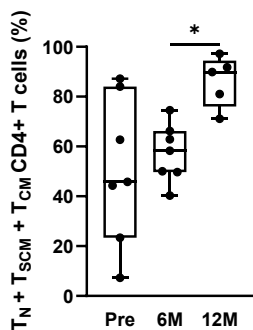
Generation process efficiency: expansion ability, viability and CAR transduction efficacy



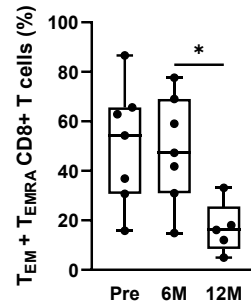
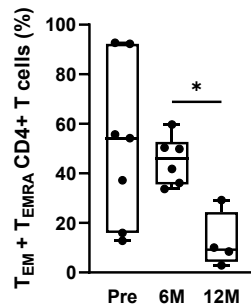
Unpublished data, international workshop on CLL (iwCLL) Congress 2025 (poster #1902)

Targeted therapy induces changes in the subpopulation frequency of CAR T cells

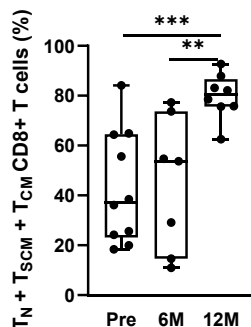
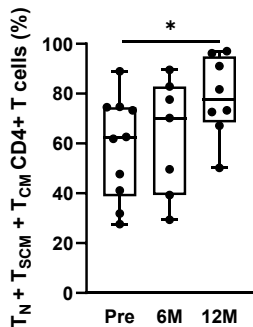
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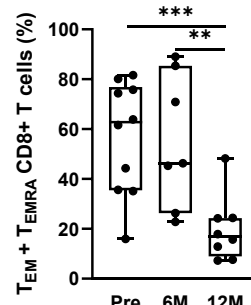
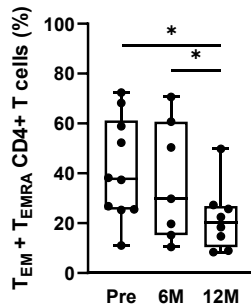
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VENETOCLAX



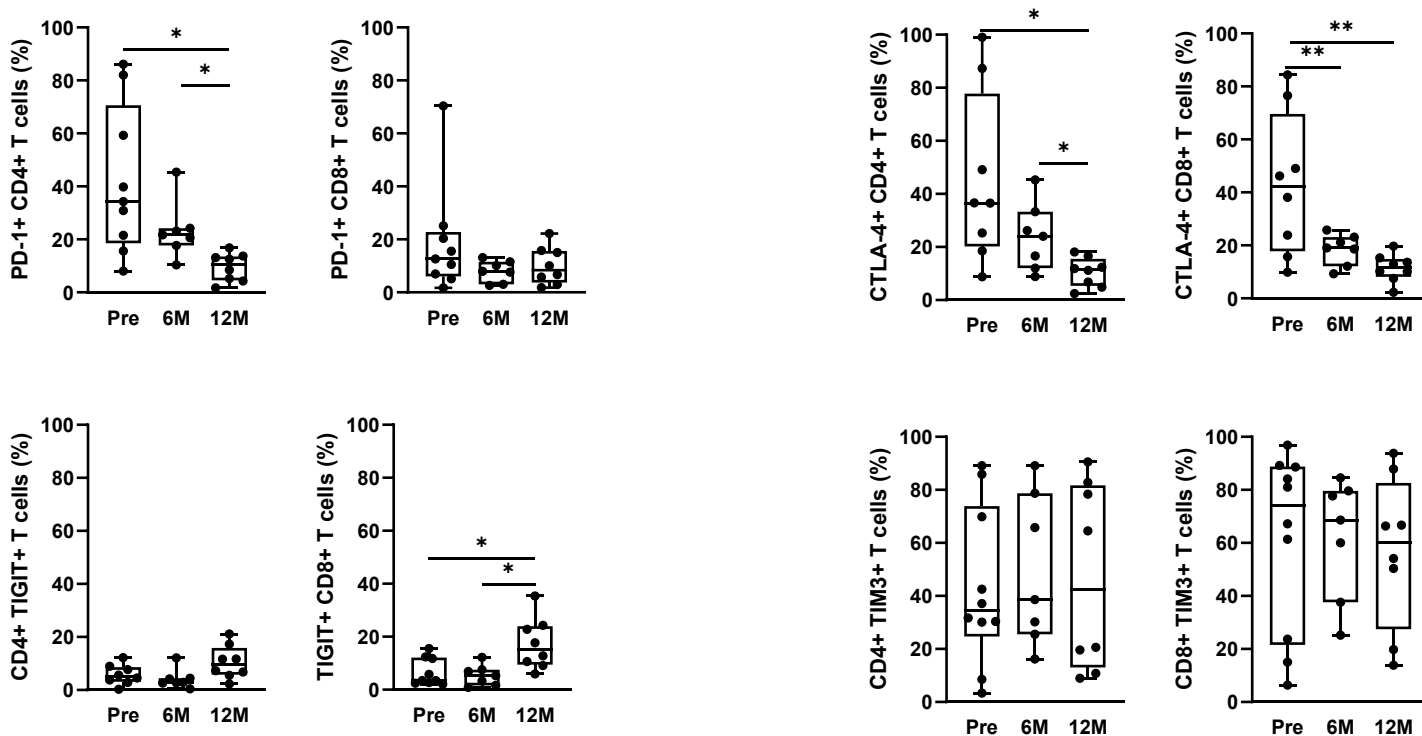
VENETOCLAX



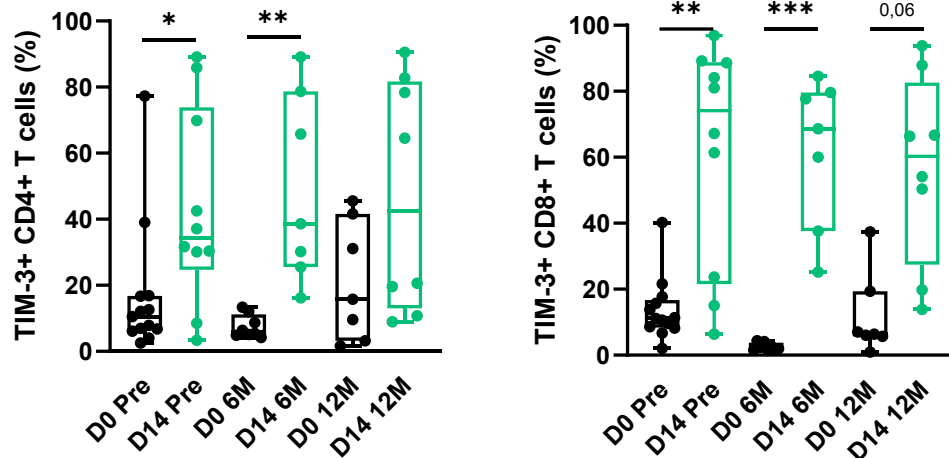
↑ Less differentiated T-cell subsets

↓ More differentiated T-cell subsets

Unpublished data, international workshop on CLL (iwCLL) Congress 2025 (poster #1902)

Treatment with **venetoclax** modulates the expression of immune checkpoint molecules

Unpublished data, international workshop on CLL (iwCLL) Congress 2025 (poster #1902)

TIM-3 expression increases during *ex vivo* CAR T-cell expansion

Prolonged *ex vivo* culture can alter T-cell fitness through progressive acquisition of exhaustion-associated phenotypes

CD19 CAR T-cell functional assessments

CAR T cells efficiently eliminate CD19+ target cells

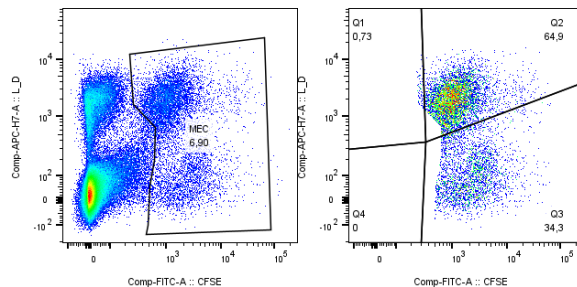
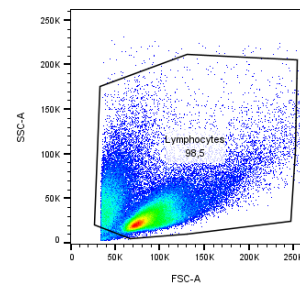
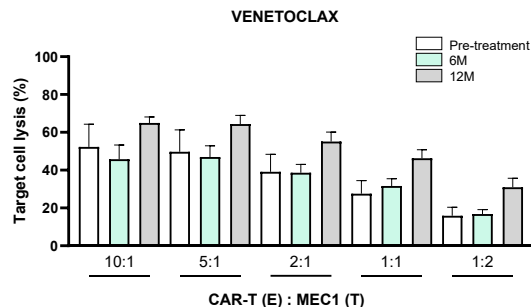
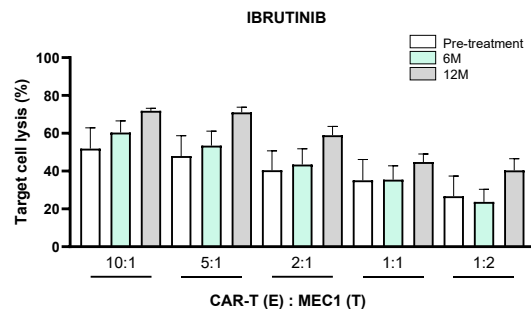
Target cell killing

Proliferation ability

Cytokine production
(i.e. IFN γ , TNF α)

Degranulation assay

Autologous CLL cells are not always available for functional assays



Conventional 2D coculture assays do not fully recapitulate the CLL tumor microenvironment

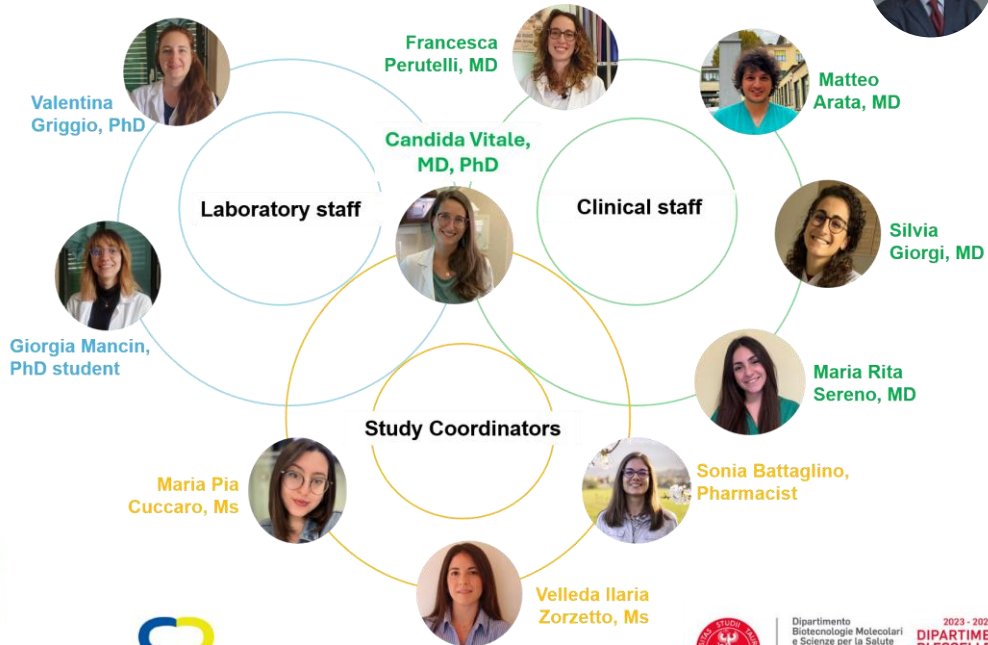
Unpublished data, international workshop on CLL (iwCLL) Congress 2025 (poster #1902)

Future applications and optimization

- An in-house small-scale manufacturing platform was successfully established for the generation and characterization of **CLL patient-derived CD19 CAR T cells**
- This platform provides a versatile tool to investigate the impact of **patient-, disease- and treatment-related variables** on CAR T-cell manufacturing and fitness, enabling direct comparisons between starting T cells and final CAR T-cell products
- The workflow can be further **optimized**, particularly by reducing manufacturing time and implementing more physiologically relevant functional models
- The platform may also serve as a preclinical system to evaluate **immunomodulatory agents and manufacturing strategies** aimed at improving CAR T-cell quality in CLL

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Lydia Scarfò
Prof. Paolo Ghia



Prof. Robin Foà



Thank you!



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